DOI:10.16781/j.CN31-2187/R.20211296

多数据库联合分析卵巢癌预后相关基因

王 静^{1△},苏晓玲^{2△},贺海威¹,王志明¹,陆 楠¹,徐明娟^{1*}
1.海军军医大学(第二军医大学)第一附属医院妇产科,上海 200433
2.海军军医大学(第二军医大学)海军特色医学中心妇产科,上海 200433

[摘要] **1** 6 寻找卵巢癌预后的关键基因,为卵巢癌治疗提供新的靶点。**方法** 从基因表达汇编(GEO)数据库 GSE18520和 GSE14407数据集、癌症基因组图谱(TCGA)数据库及基因型-组织表达(GTEx)数据库中下载 卵巢癌相关数据,用 R 3.6.2软件 limma 包进行差异表达基因分析,随后使用 R 3.6.2软件 clusterProfiler包对差异表达基因进行基因本体(GO)及京都基因与基因组百科全书(KEGG)富集分析。使用 STRING数据库建立蛋白质-蛋白质相互作用网络,利用 Cytoscape软件 cytoHubba 插件筛选核心基因,利用基因表达谱交互分析(GEPIA)数据库验证核 心基因在卵巢癌组织中的表达情况,随后使用 Kaplan-Meier Plotter数据库对核心基因进行生存分析。结果 通过 GEO数据库 GSE18520、GSE14407数据集及 TCGA、GTEx数据库共同筛选获得 69个差异表达基因,主要富集在 ABC转运体、视黄醇代谢及 Wnt 信号通路。蛋白质-蛋白质相互作用网络分析提示共有 9 个核心基因,GEPIA数据库分析结果表明这 9 个基因在卵巢癌中高表达。Kaplan-Meier Plotter数据库分析结果表明,中心体相关蛋白 55 (CEP55)、序列相似性 83家族蛋白成员 D(FAM83D)、驱动蛋白家族成员 20A (KIF20A)、细胞周期依赖性激酶亚基蛋白 2 (CKS2)和中心体相关激酶 2 (NEK2)基因高表达的卵巢癌患者总生存期比低表达的患者缩短,CEP55、FAM83D、KIF20A、CKS2、NEK2、FOXMI和 TTK 的表达与卵巢癌患者的预后密切相关。

[关键词] 卵巢肿瘤; 预后; 生物信息学; 差异表达基因

[中图分类号] R 737.31 [文献标志码] A [文章编号] 2097-1338(2022)02-0167-07

Prognostic genes in ovarian cancer: a multi-database analysis

WANG Jing¹, SU Xiao-ling², HE Hai-wei¹, WANG Zhi-ming¹, LU Nan¹, XU Ming-juan¹

1. Department of Obstetrics and Gynecology, The First Affiliated Hospital of Naval Medical University (Second Military Medical University), Shanghai 200433, China

2. Department of Obstetrics and Gynecology, Naval Medical Center, Naval Medical University (Second Military Medical University), Shanghai 200433, China

[Abstract] Objective To search for the hub genes for the prognosis of ovarian cancer and provide new targets for the treatment of ovarian cancer. Methods Ovarian cancer related data were downloaded from Gene Expression Omnibus (GEO) database (GSE18520 and GSE14407 datasets), The Cancer Genome Atlas (TCGA) database and the Genotype-Tissue Expression (GTEx) database. Differentially expressed genes were analyzed with limma package of R 3.6.2 software, and then clusterProfiler package was used for Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses of these genes. Meanwhile, STRING was used to establish the protein-protein interaction network, and cytoHubba package of Cytoscape software was used to screen the hub genes. Gene Expression Profile Interaction Analysis (GEPIA) database was used to verify the expression of hub genes in ovarian cancer tissues. Then, Kaplan-Meier Plotter database was used to perform survival analysis on the hub genes. Results A total of 69 differentially expressed genes were screened by GEO (GSE18520 and GSE14407), TCGA and GTEx databases, and they were mainly enriched in the ABC transporter, retinol metabolism and Wnt signaling pathways. Protein-protein interaction network analysis showed that there were 9 hub genes, which were verified in GEPIA. Kaplan-Meier Plotter database analysis showed that the overall survival was

[收稿日期] 2021-12-23 [接受日期] 2022-01-27

[基金项目] 海军军医大学(第二军医大学)第一附属医院"234 学科攀峰计划"(2019YXK014),深蓝 123 重点攻关项目(2020YSL009),海军计 生课题(19JSZ05). Supported by the "234 Discipline Peak Climbing Plan" of The First Affiliated Hospital of Naval Medical University (Second Military Medical University) (2019YXK014), Key Project of "Shenlan 123" (2020YSL009), and Navy Family Planning Project (19JSZ05).

[作者简介] 王 静,博士生. E-mail: 13296360601@163.com;苏晓玲,博士生. E-mail: 18806285788@163.com



[△]共同第一作者(Co-first authors).

^{*}通信作者(Corresponding author). Tel: 021-31162044, E-mail: 13634373419@163.com

shorter in the ovarian cancer patients with high expression of centrosomal protein 55 (*CEP55*), family with sequence similarity 83, member D (*FAM83D*), kinesin family member 20A (*KIF20A*), cyclin dependent-kinase subunit protein 2 (*CKS2*) and NIMA related kinase 2 (*NEK2*) genes; and the progression-free survival was shorter in patients with high expression of *CEP55*, *FAM83D*, *KIF20A*, forkhead box protein M1 (*FOXM1*) and TTK protein kinase (*TTK*) than those with low expression. **Conclusion** The expression of *CEP55*, *FAM83D*, *KIF20A*, *CKS2*, *NEK2*, *FOXM1* and *TTK* are closely related to the prognosis of ovarian cancer patients.

[Key words] ovarian neoplasms; prognosis; bioinformatics; differentially expressed genes

[Acad J Naval Med Univ, 2022, 43(2): 167-173]

卵巢癌的发病率和病死率在女性恶性肿瘤中均 排名第8位^[1]。早期缺乏有效的诊断方法与晚期 高复发率是造成卵巢癌高病死率的主要原因^[2]。 因此,寻找有效的肿瘤标志物并研究其在卵巢癌发 生、发展中的作用对卵巢癌的诊断、预防和治疗具 有重要意义。本研究通过生物信息学技术寻找与卵 巢癌预后相关的关键基因,为卵巢癌的治疗提供新 的靶点。

1 资料和方法

1.1 基因芯片数据获取 从基因表达汇编(Gene Expression Omnibus, GEO)数据库下载GSE18520 和GSE14407数据集中的相关数据,前者包括53 个肿瘤样本和10个正常卵巢样本,后者包括12 个肿瘤样本和12个正常卵巢样本,所用平台均为GPL570(Affymetrix Human Genome U133 Plus 2.0 Array)。从基因型-组织表达(Genotype-Tissue Expression,GTEx)数据库中获得88 例卵巢正常组织的数据,从癌症基因组图谱(The Cancer Genome Atlas,TCGA)数据库中获得379 例卵巢 癌组织的数据。

1.2 差异表达基因获取 使用R 3.6.2 软件 limma 包筛选卵巢癌与正常组织之间的差异表达基因,

以 *P*<0.05 和 |log₂FC|>2 [FC 为 差 异 倍 数 (fold change)]为筛选标准。使用维恩图获得 GEO 数据 库 GSE18520、GSE14407 数 据集 及 TCGA、GTEx 数据库中重叠的基因。

基因功能和通路富集分析 利用R 3.6.2 软件 clusterProfiler 包进行基因本体 (gene ontology,

GO) 和京都基因与基因组百科全书 (Kyoto Encyclopedia of Genes and Genomes, KEGG) 富集 分析, 设定 P < 0.05 为差异有统计学意义。

1.4 核心基因筛选 使用 STRING 数据库(http:// string-db.org/)进行蛋白质-蛋白质相互作用 (protein-protein interaction, PPI)网络构建。通 过 Cytoscape 软件^[3]进一步分析 PPI 网络,利用 cytoHubba插件^[4]根据Matthews 相关系数(Matthews correlation coefficient, MCC)算法筛选核心基因。 1.5 核心基因验证 利用基因表达谱交互分析 (Gene Expression Profiling Interactive Analysis, GEPIA)数据库(*http://gepia.cancer-pku.cn*)^[5]验 证获得的核心基因在卵巢癌中的表达情况。通过 人类蛋白质图谱(Human Protein Atlas, HPA)数 据库(*https://www.proteinatlas.org*)^[6]分析核心基 因在卵巢癌和正常组织中的蛋白表达。在Kaplan-Meier Plotter 数据库(*https://www.kmplot.com*)中 对核心基因进行生存分析。

2 结 果 家

2.1 差 异 表 达 基 因 分 析 通 过 GEO 数 据 库 GSE18520 和 GSE14407 数据集获得了 211 个共同 的正常卵巢样本与卵巢肿瘤组织的差异表达基因,

其中 53 个表达上调, 158 个表达下调。通过 GTEx 结合 TCGA 数据库进行分析,获得 2 253 个差异表 达基因,其中 1 017 个表达上调,1 236 个表达下调。进一步分析发现,TCGA 数据库中有 69 个差异表 达基因与 GEO 数据库 GSE18520 和 GSE14407 数 据集的重叠基因匹配,其中 35 个表达上调、34 个 表达下调。对 69 个基因进行 GO 富集分析发现, 生物过程主要富集于间充质细胞分化和泌尿生殖系 统中,细胞组分主要富集于纺锤体,分子功能主要 富集于受体 – 配体活性等;KEGG 富集分析显示,信号通路主要富集在 ABC 转运体、视黄醇代谢和 Wnt 信号通路(图1)。

2.2 核心基因获取和验证 在 STRING 中构建了 69 个差异基因的 PPI 网络,包括 26 个节点和 70 个 边,经计算获得 10 个核心基因,其中 BUB1 有丝 分裂检查点丝氨酸/苏氨酸激酶 B (BUB1 mitotic

体相关激酶 2(NIMA related kinase 2, NEK2)、 驱动蛋白家族成员 20A(kinesin family member 20A, KIF20A)、TTK蛋白激酶(TTK protein kinase, TTK)、驱动蛋白家族成员 4A(kinesin family member 4A, KIF4A)。GEPIA数据库分析 表明,这9个核心基因在卵巢癌组织中高表达 (图 2),并且 NEK2表达水平随卵巢癌分期的增 高而降低(图 3)。通过 HPA 数据库获取卵巢癌

患者的临床免疫组织化学染色样本,结果显示卵巢

癌组织中CEP55和KIF20A蛋白呈高表达(图4)。

checkpoint serine/threonine kinase B, *BUB1B*) 基因 在卵巢癌中研究颇多,故本文中不做具体分析; 其余9个基因分别为细胞周期依赖性激酶亚基蛋 白2(cyclin dependent-kinase subunit protein 2, *CKS2*)、母体胚胎亮氨酸拉链激酶(maternal embryonic leucine zipper kinase, *MELK*)、序列相 似性83家族蛋白成员D(family with sequence similarity 83, member D; *FAM83D*)、中心体相 关蛋白55(centrosomal protein 55, *CEP55*)、叉 头框蛋白M1(forkhead box M1, *FOXM1*)、中心





Fig 1 GO and KEGG enrichment analyses of differentially expressed genes of ovarian cancer

A: GO functional enrichment analysis; B: KEGG pathway enrichment analysis. GO: Gene Ontology; KEGG: Kyoto Encyclopedia of Genes and Genomes.



图 2 GEPIA 数据库中卵巢正常组织与卵巢癌组织核心基因的表达

Fig 2 Expression of hub genes in normal and ovarian cancer tissues in GEPIA database

*P < 0.05. n = 426 in ovarian cancer tissue (T) group, n = 88 in normal ovarian tissue (N) group. GEPIA: Gene Expression Profile Interaction Analysis; KIF20A: Kinesin family member 20A; TTK: TTK protein kinase; NEK2: NIMA related kinase 2; MELK: Maternal embryonic leucine zipper kinase; KIF4A: Kinesin family member 4A; FOXM1: Forkhead box M1; FAM83D: Family with sequence similarity 83, member D; CKS2: Cyclin dependent-kinase subunit protein 2; CEP55: Centrosomal protein 55.





Fig 3 Relationship between hub genes and ovarian cancer stages in GEPIA database

GEPIA: Gene Expression Profile Interaction Analysis; KIF20A: Kinesin family member 20A; NEK2: NIMA related kinase 2; FAM83D: Family with sequence similarity 83, member D; CEP55: Centrosomal protein 55.



图 4 HPA 数据库中正常卵巢组织与卵巢癌组织核心基因的蛋白表达情况

Fig 4 Protein expression of hub genes in normal ovarian tissues and ovarian cancer tissues in HPA database A: KIF20A; B: CEP55; C: NEK2. Immunohistochemistry (40×). HPA: Human Protein Atlas; KIF20A: Kinesin family member 20A; CEP55: Centrosomal protein 55; NEK2: NIMA related kinase 2.

Kaplan-Meier Plotter 数据库生存分析结果显(图 5),而 CEP55、FOXM1、FAM83D、KIF20A示, CEP55、CKS2、FAM83D、KIF20A 和 NEK2和 TTK 高表达的卵巢癌患者无进展生存期比低表高表达的卵巢癌患者总生存期比低表达的患者短达的患者短(图 6)。



图 5 Kaplan-Meier Plotter 数据库中核心基因表达与卵巢癌患者总生存期的关系

Fig 5 Association of expression of hub genes with overall survival of ovarian cancer patients in Kaplan-Meier Plotter database

KIF20A: Kinesin family member 20A; TTK: TTK protein kinase; NEK2: NIMA related kinase 2; MELK: Maternal embryonic leucine zipper kinase; KIF4A: Kinesin family member 4A; FOXM1: Forkhead box M1; FAM83D: Family with sequence similarity 83, member D; CKS2: Cyclin dependent-kinase subunit protein 2; CEP55: Centrosomal protein 55; *HR*: Hazard ratio; *CI*: Confidence interval.



图 6 Kaplan-Meier Plotter 数据库中核心基因表达与卵巢癌患者无进展生存期的关系 Fig 6 Association of expression of hub genes with progress free survival of ovarian cancer patients in Kaplan-Meier

Plotter database

KIF20A: Kinesin family member 20A; TTK: TTK protein kinase; NEK2: NIMA related kinase 2; MELK: Maternal embryonic leucine zipper kinase; KIF4A: Kinesin family member 4A; FOXM1: Forkhead box M1; FAM83D: Family with sequence similarity 83, member D; CKS2: Cyclin dependent-kinase subunit protein 2; CEP55: Centrosomal protein 55; *HR*: Hazard ratio; *CI*: Confidence interval.

3 讨 论

虽然卵巢癌的治疗方法和手术方式已经有所 改进,但晚期卵巢癌患者由于诊断困难,治疗结果 和预后仍然很差,探索与卵巢癌预后相关的基因非 常必要。本研究从卵巢癌组织和正常卵巢组织芯 片数据中得到 69 个差异表达基因 (其中上调基因 35个,下调基因34个)。GO 富集分析显示,在生 物学过程中, 差异基因主要集中在间充质细胞分化 及泌尿生殖系统中。大量研究表明上皮-间质转化 (epithelial-mesenchymal transition, EMT) 在胚胎 发育中发挥了关键作用,同时也参与了肿瘤的进展 和转移^[7-9];上皮钙黏蛋白(epithelial cadherin, E-cadherin) 与神经钙黏蛋白 (neural cadherin, N-cadherin)是EMT中的重要分子,研究发现, E-cadherin 表达增多与 N-cadherin 表达下降可降低 卵巢癌细胞的侵袭能力^[9]。KEGG 富集分析发现, 差异表达基因主要富集在 ABC 转运体、视黄醇代 谢和 Wnt 信号通路。卵巢癌是一种常见的易出现 化学治疗耐药的实体肿瘤,既往研究发现ABC转 运蛋白可致癌症的多药耐药,包括多柔比星、依托 泊苷和长春新碱等^[10],而ABC转运蛋白在卵巢癌 中的研究并不多见,这也为研究卵巢癌化学治疗耐 药提供了新的思路。视黄醇代谢已被证明与乳腺 癌和胆囊癌有关^[11]。在胚胎和成人组织稳态中, Wnt/β-catenin 通路调节细胞增殖、极性、存活和干 细胞命运, Wnt 信号通路异常与肿瘤的发生等多种 病理过程有关^[12-13],越来越多的研究证明 Wnt 信 号通路影响卵巢癌的血管生成、转移、化学治疗耐 药和免疫逃逸等诸多方面^[14-15]。

本研究通过PPI网络分析筛选出9个核心基 因。在卵巢癌组织中这9个核心基因的表达均高 于卵巢正常组织。生存分析结果显示, CEP55、 CKS2、FAM83D、KIF20A 和 NEK2 高表达的患者总 生存期较短,并且其中CEP55、FAM83D和KIF20A 也与患者的无进展生存期有关。此外, NEK2与 卵巢癌分期相关。近年研究发现 CEP55 参与调控 PI3K/AKT 通路和癌细胞干细胞化^[16-18]。临床研究 发现 CEP55 在乳腺癌、前列腺癌、肾癌、甲状腺 癌等多种癌症中高表达^[19-20],高表达的 CEP55 蛋 白与非小细胞肺癌的不良预后相关^[21]。本研究发 现, CEP55 高表达的卵巢癌患者的总生存期和无进 展生存期比低表达的患者短。CKS2属于细胞周期 依赖蛋白激酶亚基家族,参与细胞周期调控^[22]。 研究表明, CKS2 表达下调可抑制结直肠癌患者肿 瘤细胞增殖、促进凋亡^[23]。本研究发现 CKS2 的 高表达预示着卵巢癌患者总生存期较差,但不影响 患者的无进展生存期。FAM83D可能通过抑制抑 癌因子含 F 框和 WD 重复域蛋白 7 (F-box and WD repeat domain containing 7, FBXW7) 在乳腺癌中 发挥致癌作用^[24],同时可以促进肝癌增殖和侵 袭^[25]。本研究结果表明, FAM83D 高表达的卵巢 癌患者总生存期和无进展生存期较差。KIF20A与 细胞增殖、迁移和化学治疗耐药有关。许多研究证 实, KIF20A 在肺癌^[26]、胃癌^[27]、肝癌^[28]等恶性 肿瘤中高表达, 然而, 其与卵巢癌的相关性尚不清 楚。在本研究中, KIF20A 高表达卵巢癌患者总生 存期和无进展生存期较差。NEK2 是宫颈癌组织中 过表达的丝氨酸/苏氨酸激酶,与肿瘤分期和淋巴 结转移有关^[29]。本研究发现 NEK2 与卵巢癌的预 后和卵巢癌的肿瘤分期有关,同时其高表达也与卵 巢癌患者预后不良有关。以上这些基因与不同癌症 的发生、发展密切相关,但在卵巢癌中的研究并不 多见,因其与卵巢癌的预后密切相关,因此后期进 行体内及体外实验的验证是非常必要的。

本研究结果显示, CEP55、CKS2、FAM83D、 KIF20A和NEK2在卵巢癌组织中的mRNA水平高 于正常卵巢组织;但根据HPA结果,只有CEP55 和KIF20A的蛋白水平在肿瘤组织中高于正常组 织;这可能与蛋白质的某些修饰有关,但具体机制 尚不清楚。

综上所述,本研究通过对多数据库进行分析, 发现*CEP55、CKS2、FAM83D、KIF20A、NEK2、 FOXM1、TTK*与卵巢癌患者预后相关;此外, *NEK2*与卵巢癌分期相关。本研究仅为基于多数据 库的分析结果,因此后期需要从细胞与动物实验方 面进行以上基因的验证,其具体作用机制也有待进 一步探索。

[参考文献]

- [1] SUNG H, FERLAY J, SIEGEL R L, LAVERSANNE M, SOERJOMATARAM I, JEMAL A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries[J]. CA Cancer J Clin, 2021, 71: 209-249.
- [2] XU Z Z, ZHOU Y, CAO Y X, DINH T L A, WAN J, ZHAO M. Identification of candidate biomarkers and analysis of prognostic values in ovarian cancer by integrated bioinformatics analysis[J/OL]. Med Oncol, 2016, 33: 130. DOI: 10.1007/s12032-016-0840-y.
- [3] SU G, MORRIS J H, DEMCHAK B, BADER G D. Biological network exploration with Cytoscape 3[J/OL].

Curr Protoc Bioinformatics, 2014, 47: 8.13.1-8.1324. DOI: 10.1002/0471250953.bi0813s47.

- [4] CHIN C H, CHEN S H, WU H H, HO C W, KO M T, LIN C Y. cytoHubba: identifying hub objects and subnetworks from complex interactome[J/OL]. BMC Syst Biol, 2014, 8: S11. DOI: 10.1186/1752-0509-8-S4-S11.
- [5] TANG Z F, LI C W, KANG B X, GAO G, LI C, ZHANG Z M. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses[J/OL]. Nucleic Acids Res, 2017, 45: W98-W102. DOI: 10.1093/nar/gkx247.
- [6] UHLEN M, ZHANG C, LEE S, SJÖSTEDT E, FAGERBERG L, BIDKHORI G, et al. A pathology atlas of the human cancer transcriptome[J/OL]. Science, 2017, 357: eaan2507. DOI: 10.1126/science.aan2507.
- [7] DUAN H Y, YAN Z Q, CHEN W, WU Y, HAN J S, GUO H Y, et al. TET1 inhibits EMT of ovarian cancer cells through activating Wnt/β-catenin signaling inhibitors DKK1 and SFRP2[J]. Gynecol Oncol, 2017, 147: 408-417.
- [8] IWATSUKI M, MIMORI K, YOKOBORI T, ISHI H, BEPPU T, NAKAMORI S, et al. Epithelialmesenchymal transition in cancer development and its clinical significance[J]. Cancer Sci, 2010, 101: 293-299.
 [9] ROSSO M, MAJEM B, DEVIS L, LAPYCKYJ L, BESSO M J, LLAURADÓ M, et al. E-cadherin: a determinant molecule associated with ovarian cancer progression, dissemination and aggressiveness[J/OL]. PLoS One, 2017, 12: e0184439. DOI: 10.1371/journal. pone.0184439.
- [10] AMAWI H, SIM H M, TIWARI A K, AMBUDKAR S V, SHUKLA S. ABC transporter-mediated multidrugresistant cancer[J]. Adv Exp Med Biol, 2019, 1141: 549-580.
 - [11] CHEN A C, GUO X, DERGUINI F, GUDAS L J. Human breast cancer cells and normal mammary epithelial cells: retinol metabolism and growth inhibition by the retinol metabolite 4-oxoretinol[J]. Cancer Res, 1997, 57: 4642-4651.
 - [12] CLEVERS H. Wnt/β-catenin signaling in development and disease[J]. Cell, 2006, 127: 469-480.
 - [13] POLAKIS P. The many ways of Wnt in cancer[J]. Curr Opin Genet Dev, 2007, 17: 45-51.
 - [14] AREND R C, LONDOÑO-JOSHI A I, STRAUGHN J M Jr, BUCHSBAUM D J. The Wnt/β-catenin pathway in ovarian cancer: a review[J]. Gynecol Oncol, 2013, 131: 772-779.
 - [15] NAGARAJ A B, JOSEPH P, KOVALENKO O, SINGH S, ARMSTRONG A, REDLINE R, et al. Critical role of Wnt/β-catenin signaling in driving epithelial ovarian cancer platinum resistance[J]. Oncotarget, 2015, 6:

23720-23734.

- [16] CHEN C H, LU P J, CHEN Y C, FU S L, WU K J, TSOU A P, et al. FLJ10540-elicited cell transformation is through the activation of PI3-kinase/AKT pathway[J]. Oncogene, 2007, 26: 4272-4283.
- [17] WANG G Z, LIU M N, WANG H J, YU S, JIANG Z F, SUN J H, et al. Centrosomal protein of 55 regulates glucose metabolism, proliferation and apoptosis of glioma cells via the Akt/mTOR signaling pathway[J]. J Cancer, 2016, 7: 1431-1440.
- [18] KUO T C, CHEN C T, BARON D, ONDER T T, LOEWER S, ALMEIDA S, et al. Midbody accumulation through evasion of autophagy contributes to cellular reprogramming and tumorigenicity[J]. Nat Cell Biol, 2011, 13: 1214-1223.
- [19] SHIRAISHI T, TERADA N, ZENG Y, SUYAMA T, LUO J, TROCK B, et al. Cancer/testis antigens as potential predictors of biochemical recurrence of prostate cancer following radical prostatectomy[J/OL]. J Transl Med, 2011, 9: 153. DOI: 10.1186/1479-5876-9-153.
- [20] JONES J, OTU H, SPENTZOS D, KOLIA S, INAN M, BEECKEN W D, et al. Gene signatures of progression and metastasis in renal cell cancer[J]. Clin Cancer Res, 2005, 11: 5730-5739.
- [21] JIANG C, ZHANG Y, LI Y, LU J, HUANG Q, XU R, et al. High CEP55 expression is associated with poor prognosis in non-small-cell lung cancer[J]. Onco Targets Ther, 2018, 11: 4979-4990.
- [22] CHEN R, FENG C, XU Y. Cyclin-dependent kinaseassociated protein CKS2 is associated with bladder cancer progression[J]. J Int Med Res, 2011, 39: 533-540.
- [23] YU M H, LUO Y, QIN S L, WANG Z S, MU Y F, ZHONG M. Up-regulated CKS2 promotes tumor

progression and predicts a poor prognosis in human colorectal cancer[J]. Am J Cancer Res, 2015, 5: 2708-2718.

- [24] WANG Z R, LIU Y Y, ZHANG P J, ZHANG W G, WANG W J, CURR K, et al. FAM83D promotes cell proliferation and motility by downregulating tumor suppressor gene *FBXW7*[J]. Oncotarget, 2013, 4: 2476-2486.
- [25] LIAO W J, LIU W L, LIU X, YUAN Q, OU Y, QI Y, et al. Upregulation of FAM83D affects the proliferation and invasion of hepatocellular carcinoma[J]. Oncotarget, 2015, 6: 24132-24147.
- [26] ZHAO X, ZHOU L L, LI X Y, NI J, CHEN P, MA R, et al. Overexpression of KIF20A confers malignant phenotype of lung adenocarcinoma by promoting cell proliferation and inhibiting apoptosis[J]. Cancer Med, 2018, 7: 4678-4689.
- [27] YAN G R, ZOU F Y, DANG B L, ZHANG Y, YU G C, LIU X, et al. Genistein-induced mitotic arrest of gastric cancer cells by downregulating KIF20A, a proteomics study[J]. Proteomics, 2012, 12: 2391-2399.
- [28] GASNEREAU I, BOISSAN M, MARGALL-DUCOS G, COUCHY G, WENDUM D, BOURGAIN-GUGLIELMETTI F, et al. KIF20A mRNA and its product MKlp2 are increased during hepatocyte proliferation and hepatocarcinogenesis[J]. Am J Pathol, 2012, 180: 131-140.
- [29] XU T, ZENG Y, SHI L, YANG Q, CHEN Y, WU G, et al. Targeting NEK2 impairs oncogenesis and radioresistance via inhibiting the Wnt1/β-catenin signaling pathway in cervical cancer[J/OL]. J Exp Clin Cancer Res, 2020, 39: 183. DOI: 10.1186/s13046-020-01659-y.

[本文编辑] 孙 岩